US-SOMO (SOlution MOdeler): versatile and reliable hydrodynamic and SAS modeling of biomacromolecules within the UltraScan AUC data analysis software

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Hydrodynamics. Stokes-Einstein's law The translational frictional coefficient of **f** a sphere of

radius **r** in a solvent of viscosity η is:

$f = 6\pi\eta r$

For a body of unknown shape having a translational frictional coefficient f, we can define the Stokes radius R_s as that of a sphere having the same f

$$R_s = r = f/6\pi\eta$$

Translational diffusion coefficient D_t : $D_t = k_b T/f$ where $k_b = Boltzmann's constant$ T = solution temperature (°K)

Sedimentation coefficient s:

 $s = (M/N_0)(1 - V_2\rho)/f$

where M = mass $N_0 = Avogadro's number$ $v_2 = partial specific volume$ $\rho = solvent density$

Early applications of hydrodynamics: representing proteins as simple geometrical objects (spheres, ellipsoids of revolution)



FIGURE 7-25. Dimensions of the ellipsoid of revolution that best account for the hydrodynamic properties (viscosity and frictional coefficient) of various protein molecules. [After W. J. Moore, *Physical Chemistry*, Prentice-Hall, Englewood Cliffs, N. J., 1972.)

Early applications of hydrodynamics: representing proteins as simple geometrical objects (spheres, ellipsoids of revolution)



Cantor & Schimmel, Biophysical Chemistry Part II: Techniques for the study of biological strcuture and function

The "hydration" issue

Cantor and Schimmel - Biophysical Chemistry Part II, 1980

12-1 VISCOMETRY 655

Table 12-2

Hydrations of biopolymers computed by using shapes known from x-ray diffraction or electron microscopy

			Hydration (δ_1 in g/g) based on							
Sample	Known axial ratio [§]	in the second	Viscosity			Diffusion S			Sedimentation	
Bushy stunt virus	1.0	5	0.65			0.71	N. S. W.	Alexandri	0.71	net).
Carboxypeptidase	1.25		<u> </u>			0.30			0.69	
Cytochrome c	1.48					0.18			0.24	
Hemoglobin	1.3		0.62			0.52			0.75	
Lysozyme	1.5		0.34			0.52			0.52	
Myoglobin	1.76		0.44			0.50			0.42	
Tobacco mosaic virus	18		0.32			0.1 - 0.7			0.26	

[§]Axial ratios are for prolate ellipsoids, except for cytochrome c, which is oblate.

SOURCE: After I. D. Kuntz, Jr., and W. Kauzmann, in Advances in Protein Chemistry, vol. 28, ed. C. B. Anfinsen, J. T. Edsall, and F. M. Richards (New York: Academic Press, 1974), p. 239.

Advances in Protein Chemistry 28:239-345, 1974

HYDRATION OF PROTEINS AND POLYPEPTIDES

By I. D. KUNTZ, JR. and W. KAUZMANN

Department of Pharmaceutical Chemistry, University of California, San Francisco, California, and Department of Chemistry, Princeton University, Princeton, New Jersey

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Hydration from NMR freezing experiments

I. D. KUNTZ, JR. AND W. KAUZMANN

HYDRATION OF PROTEINS AND POLYPEPTIDES

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TABLE XXII

Proposed Amino Acid Hydrations Based on Nuclear Magnetic Resonance Studies of Polypeptides^a

In agricult Resonance Statuces of I bigpeptiace

Amino acid residues ^b	Hydration
Ionic	
Asp ⁻	6
Glu-	7
Tyr-	7
Arg ⁺	3
His ⁺	4 ^d
Lys^+	4
Polar	
Asn	2
Gln	- 2°
Pro	3
Ser, Thr	2.
Trp	2*
Asp	2
Glu	2
Tyr	(3)
Arg	3
Lvs	4
Nonpolar	
Ala	1
Gly	1
Phe	(0)
Val	1
Ile, Leu, Met	10

^a After Kuntz (1971a).

^b Standard three-letter code.

^c Moles of water per mole of amino acid.

^d As Lys⁺.

• Assumed values based on one water molecule per amide plus one water molecule per side-chain polar group.

	TABLE	XXIII	
Prediction of Protein	Hydration from	Composition	and Polypeptide Results ^a

Hydration (g H	ion (g H_2O/g protein)		
Calculated ^b	Observed ^e		
0.36	0.34		
0.45	0.42		
0.39	0.34		
0.36	0.33		
0.37	0.33		
0.45	0.40		
0.42	0.42		
0.45	0.44		
0.32ª	0.30		
	Hydration (g F Calculated ^b 0.36 0.45 0.39 0.36 0.37 0.45 0.42 0.45 0.42 0.45 0.32 ^d		

^a After Kuntz (1971a); see Table XXII.

^b Calculation assumes that *all* residues are fully hydrated. This is perhaps reasonable for the denatured proteins but leads to a small positive error unless allowance is made for "buried" groups. This correction was done for lysozyme, yielding a calculated value of 0.335.

^c NMR freezing experiments.

^d Calculation assumes that all carboxyl groups are uncharged at pH 3.

Reconciling hydration dynamics with hydrodynamics

PNAS 100:12135-12140, 2003

Biomolecular hydration: From water dynamics to hydrodynamics

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1-The static picture of biomolecular hydration is fundamentally inconsistent with magnetic relaxation dispersion experiments and molecular dynamics simulations, which both reveal a highly dynamic interface where rotation and exchange of nearly all water molecules are several orders of magnitude faster than biomolecular diffusion.

2-Waters near the biomolecular surface have a different density, and alter the local viscosity. It turns out that considering a number of "tightly bound", static water molecules compensate well for this local viscosity effect, otherwise very hard to be directly taken into account. Bead modeling methods: from an idea of V. Bloomfield, further developed by D.C. Teller, to HYDROPRO (J. García de la Torre)









FIGURE 1 Two-dimensional analogies of the various model types. (A) A bead model (in strict sense). (B) Shell model. (C) Filling model. (D) Rough-shell model.

Carrasco & García de la Torre Bioph. J. 75, 3044-3057, 1999





FIGURE 2 (A) A bead-per-atom (BPA) model of lysozyme, which we take as the primary hydrodynamic particle (PHP) that represents this protein. The atomic element radius (AER) is a = 3 Å. (B) A shell model (SHE), derived from the PHP, used for hydrodynamic calculations. The radius of the small beads in this case is $\sigma = 0.8$ Å.

García de la Torre, Huertas & Carrasco Bioph. J. 78, 719-730, 2000

Low Reynolds number hydrodynamics

$$\begin{pmatrix} \mathbf{F} \\ \mathbf{T}_{\mathbf{O}} \end{pmatrix} = \begin{pmatrix} \mathbf{\Xi}_{\mathbf{t}} & \mathbf{\Xi}_{\mathbf{O},\mathbf{c}}^{\mathbf{T}} \\ \mathbf{\Xi}_{\mathbf{O},\mathbf{c}} & \mathbf{\Xi}_{\mathbf{O},\mathbf{r}} \end{pmatrix} \begin{pmatrix} \mathbf{u}_{\mathbf{O}} \\ \boldsymbol{\omega} \end{pmatrix}$$

 $\mathbf{F}_{i} \left(6\pi \eta_{0} \sigma_{i} \right)^{-1} + \sum_{j=1}^{N} \mathbf{T}_{ij} \cdot \mathbf{F}_{j} = (\mathbf{u}_{i} - \mathbf{v}_{i}^{0})$

$$\mathbf{T}_{ij} = (8\pi \eta_0 \mathbf{R}_{ij})^{-1} \left[\mathbf{I} + \frac{\mathbf{R}_{ij} \mathbf{R}_{ij}}{\mathbf{R}_{ij}^2} + \frac{(\sigma_i^2 + \sigma_j^2)}{\mathbf{R}_{ij}^2} \left(\frac{1}{3} \mathbf{I} - \frac{\mathbf{R}_{ij} \mathbf{R}_{ij}}{\mathbf{R}_{ij}^2} \right) \right]$$

$$\sum_{j=1}^{N} \mathbf{B}_{ij} \cdot \mathbf{F}_{j} = (\mathbf{u}_{i} - \mathbf{v}_{i}^{0}) \qquad \mathbf{B}_{ij} = \delta_{ij} \frac{\mathbf{I}}{6\pi \eta_{0} \sigma_{i}} + (1 - \delta_{ij}) \mathbf{T}_{ij} \qquad C = \mathcal{B}^{-1}$$

$$\Xi_t = \sum_i \sum_j \mathbf{C}_{ij} = \Xi_{O,c} = \sum_i \sum_j \mathbf{U}_i \cdot \mathbf{C}_{ij} = \Xi_{O,r} = -\sum_i \sum_j \mathbf{U}_i \cdot \mathbf{C}_{ij} \cdot \mathbf{U}_j + 6 \eta_0 \mathbf{V} \mathbf{I}$$

TO COMPUTE THE PARAMETERS THAT **CAN BE MEASURED EXPERIMENTALLY, A COMPROMISE MUST BE REACHED BETWEEN A GOOD REPRESENTATION OF** THE SURFACE OF THE PROTEIN AND A LOW NUMBER OF FRICTIONAL ELEMENTS (BEADS).

THE LAYER OF "TIGHTLY BOUND" WATER OF HYDRATION MUST ALSO BE TAKEN INTO ACCOUNT Early programs developed by the Byron/Rocco groups:

BEAMS (BEAds Modeling System) Spotorno et al., Eur. Biophys. J. 25, 373-384, 1997.

AtoB Byron, Biophys J. 72, 408-415, 1997.

SOMO (SOlution MOdeler) Rai et al., Structure 13, 723-744, 2005;



Method SOMO (SOlution MOdeller): generating mediumresolution bead models from atomic coordinates



Main features:

1 bead/side chain & 1 bead/main chain. Water of hydration, based on residues, is included in each bead.

$A \rightarrow B$ After ASA screening, exposed side -chains beads are placed.

 $B \rightarrow C$ Beads overlapping by more than a preset threshold can be fused together. Overlaps are then removed, reducing the radii and outward translating the centers of exposed beads.

 $C \rightarrow D$ Exposed peptide bond beads are placed and overlaps removed.

 $D \rightarrow E$ Buried beads are placed and overlaps removed. They should be excluded from the computations of the hydrodynamic parameters.

Rai et al., Structure, May 2005

The "peptide bond" rule



The "peptide bond" rule



Improved AtoB (Grid) method: generating variable-resolution bead models from atomic coordinates



Main features:

1 bead/cube in a variable-size cubic grid. Water of hydration, based on atom values, included in each bead.

 $A \rightarrow B$ All the beads are generated and placed (CM or CC).

B→C Beads are screened for surface accessibility (ASA; red, accessible; orange, buried).

 $C \rightarrow D$ Overlaps between the exposed beads are then removed, reducing the radii and outward translating the beads' centers.

 $D \rightarrow E$ Overlaps between the buried beads are then removed, and they are re-screened for accessibility. Buried beads are excluded from hydrodynamic computations.

Eur Biophys J DOI 10.1007/s00249-009-0418-0

ORIGINAL PAPER

The implementation of SOMO (SOlution MOdeller) in the UltraScan analytical ultracentrifugation data analysis suite: enhanced capabilities allow the reliable hydrodynamic modeling of virtually any kind of biomacromolecule

Emre Brookes · Borries Demeler · Camillo Rosano · Mattia Rocco

mabi.200900474

Developments in the US-SOMO Bead Modeling Suite: New Features in the Direct Residue-to-Bead Method, Improved Grid Routines, and Influence of Accessible Surface Area Screening^a

Emre Brookes, Borries Demeler, Mattia Rocco*

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Layout and features of US-SOMO, integrated in UltraScan and available for Linux, Win, and Mac							
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Add/Edit <u>A</u> f	tom	,	ZENO calculation start Calculate hydrodynamics (Zeno) completed Visualizing model 1 Peptide Bond Rule is on for this PDB				
Add/Edit <u>R</u>	esidue	lt	All options set to default values 8RAT models selected: 1				
Add/Edit <u>S</u>	AXS coeffi	cients	Building the bead model for 8RAT model 1 Checking the pdb structure PDB structure ok				
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SOMO Solution Modeler	
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1: Define Resid	lue Properties:
Load Atom Definition File	/usr/local/ultrascan/etc/somo.atom
Load Residue Definition File	/usr/local/ultrascan/etc/somo.residue
Residue Name:	TYR
Description:	Tyrosine
Number of Atoms in Residue:	¥ × 12 • ± ±
Number of Beads for Residue:	₹ ₹ 7 2
Residue Type:	Amino Acid
Residue anhydrous mol. vol. (A^3):	197.00
Residue partial spec. vol. (cm^3/g):	0.708
Max. Accessible Surface Area (A^2):	228.00
Number of Residues in File: 74	11: Amino Acid, HIS (Histidine)
Accept Residue and Continue	12: Amino Acid, PHE (Phenylalanine
	Amino Acid, LYK (Tyrosine)
Help Config 100%	

Layout and features of US-SOMO, integrated in UltraScan and available for Linux, Win, and Mac					
🎄 SOMO Solution Modeler					
Lookup Tables SOMO MD PDB	Configuration		File 1 will be included		
	2. Define R	esidue Ato	oms:		
Select Residue Atom	to be defined	Atom 1: N	(N3H1, Positioning: no) 👻		
Select Atom from Lo	okup Table:	Ν	-		
Select Hybridization	for Atom:	N3H1	▼		
Atom determines Position:		🗖 (Check i	f true)		
Hydration Number fo	or Atom:	₹ ₹ • 1			
Assign Current Atom			Continue		
Build SoMo Bead Model	Build AtoB (Gri	d) Bead Model	Begin popping stage 3 Beads popped 0.		
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The second se			Calculate budged uppmice completed		
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SOMO Solution Modeler		
Lookup Tables SOMO MD PDB Config	guration File 1 will be inc	<u>lu</u> ded
3. Define Resid	ue Bead Properties:	art amics (Zeno) completed
Select Residue Bead to be defined:	Bead 1: defined 🔹	i on for this PDB
Select Bead Color:	Blue (1)	fault values
Select Positioning Method:	Center of Gravity	≥d: 1
This Bead is part of the:	Backbone C Sidechain	In the second se
Currently defined Atoms for Bead:	Select Atom for Bead (multi-selection OK):	s in 2 chain(s) in this model 1 atomic model
Atom 1: N (N3H1, Positioning: no) Atom 2: CA (C4H1, Positioning: no) Atom 3: C (C3H0, Positioning: yes) Atom 4: O (O1H0, Positioning: yes)	Atom 1: N (N3H1, Positioning: no) Atom 2: CA (C4H1, Positioning: no) Atom 3: C (C3H0, Positioning: yes) Atom 4: O (O1H0, Positioning: yes) Atom 5: CB (C4H2, Positioning: no) Atom 6: CG (C3H0, Positioning: no) Atom 7: CD1 (C3H1, Positioning: no) Atom 8: CD2 (C3H1, Positioning: no) Atom 9: CE1 (C3H1, Positioning: yes) Atom 10: CE2 (C3H0, Positioning: yes)	ASAB1 ting ASA 16480.33 A^3 s in this model before popping 1 n stage 1 2 n stage 2 3 n stage 3 ig and radial reduction beads are exposed by rechecking beads
Bead Volume:	64.90	npleted
Bead Mol. Weight:	56.05	Fault values
Bead Hydration from Atoms' Values:	1.000000	ied
Override Bead Hydration Value:		pead count 246 vbar 0.71
Bead hydrated Volume, Radius:	88.94 A^3, 2.77 A	he matrix In Cycle 1 of 3
Accept Bead Definition	Reset	in Cycle 2 of 3 in Cycle 3 of 3 amics completed
Add Residue to File	Delete Residue	
Help	Close	

Layo	Layout and features of US-SOMO, integrated in UltraScan and available for Linux, Win, and Mac						
<u>A</u> SA (Calculatio	on					
<u>S</u> oMo	Overlap	Reduction		File 1 will be included ZENO calculation start Calculate hydrodynamics (Zeno) completed Visualizing model 1			
AtoB	(Grid) <u>O</u>	verlap Redu	iction	All options set to default values			
<u>H</u> ydro	odynami	c Calculatio	ns	BRAT models selected: 1 Building the bead model for 8RAT model 1			
<u>M</u> isce	llaneous	Options		Checking the pdb structure PDB structure ok There are 951 atoms in 2 chain(s) in this model			
<u>B</u> ead Model Output			Creating beads from atomic model Computing ASA via ASAB1 Return from Computing ASA Anhydrous volume 16480.33 A^3 There are 246 beads in this model before popping				
<u>G</u> rid Functions (AtoB)			Begin popping stage 1 Beads popped 0. Begin radial reduction stage 1				
SA <u>X</u> S	Options			Begin popping stage 2 Beads popped 0. Begin radial reduction stage 2			
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Help	Config	10)%				

Layout and features of US-SOMO, integrated in UltraScan and available for Linux, Win, and Mac X - 0 SOMO Solution Modeler Lookup Tables SOMO MD File 1 will he included PDB Configuration SOMO Accessible Surface Area Options start vnamics (Zeno) completed e is on for this PDB Accessible Surface Area Options: default values Re-check bead ASA Perform ASA Calculation Y. ected: 1 model for 8RAT model 1 -ASA Method: – structure Voronoi Tesselation (Surfrace, Tsodikov et al.) oms in 2 chain(s) in this model rom atomic model ia ASAB1 Rolling Sphere (ASAB1, Lee & Richards' Method) puting ASA e 16480.33 A^3 ads in this model before popping age 1 ASA Probe Radius (A): 1.4ction stage 1 age 2 ASA Probe Recheck Radius (A): 1.4 Ŧ ction stage 2 age 3 SOMO ASA Threshold (A^2): Ť 20ction stage 3 ping and radial reduction SOMO Bead ASA Threshold %: Ì Ţ 50 ed beads are exposed by rechecking Ŧ ng beads completed Grid ASA Threshold (A^2): 10 default values mic calculations Grid Bead ASA Threshold %: 30 duded ASAB1 Step Size (A): 1 bead count 246 vbar 0.71 or the matrix rsion Cycle 1 of 3 rsion Cycle 2 of 3 Help Close rsion Cycle 3 of 3 amics completed Help 100% Config

秦 SOMO Solution Modeler				
Lookup Tables SOMO MD PDB Conf	figuration	File 1 will be included	^	
🗙 SoMo Bead Overlap Reduc	tion Options			
SoMo Bead C	Overlap Reduction Op	ptions:		
Bead Overlap Tolerance: 🝹	5 v 0.001			
Exposed Side chain beads Exp	posed Main and side chain bead	ds Buried beads		
Overlap reduction b	etween exposed side	e chain beads is model		
Fuse Beads that overlap b	by more than:	₹ ₹ ▼ 70 ▲ ★ ★		
Remove Overlaps		re popping		
		Overlap Reduction Step Size (in %):		
Remove Overlaps synchronously:				
Remove Overlaps hierarchically (larger -> smaller)		▼ 1 ▲ ▲ by rechecking		
Outward Translation				
Help	Close		E	
Calculate RB Hydrodynamics SMI Ca	alculate RB Hydrodynamics SMI Calculate RB Hydrodynamics ZENO Processing model 1 bead count 246			
Show Hydrodynamic Calculations Open Hydrodynamic Calculations File		Using 98 beads for the matrix Supermatrix inversion Cycle 1 of 3		
Select Parameters to be Saved Save parameters to file		Supermatrix inversion Cycle 2 of 3 Supermatrix inversion Cycle 3 of 3 Calculate hydrodynamics completed		
BEST Model classifier	Stop Close		T	
Help Config	100%			

Layout and features of US-SOMO, integrated in UltraScan and available for Linux, Win, and Mac - O X SOMO Solution Modeler Lookup Tables SOMO MD PDB Configuration File 1 will be included _ _ × eted Grid Bead Overlap Reduction Options Grid Bead Overlap Reduction Options: Bead Overlap Tolerance: 💆 🗸 0.001 놀 Exposed grid beads Buried grid beads Non-screened grid beads el 1 Overlap reduction between exposed grid beads is model Fuse Beads that overlap by more than: ž 최훞 0 bre popping 5 Remove Overlaps Overlap Reduction Step Size (in %): Remove Overlaps synchronously: * tion Ŧ Remove Overlaps hierarchically (larger -> smaller) 2 \$ by rechecking Outward Translation Close Help Calculate RB Hydrodynamics SMI Calculate RB Hydrodynamics ZENO Processing model 1 bead count 246 vbar 0.71 Using 98 beads for the matrix Show Hydrodynamic Calculations Open Hydrodynamic Calculations File Supermatrix inversion Cycle 1 of 3 Supermatrix inversion Cycle 2 of 3 Select Parameters to be Saved Save parameters to file Supermatrix inversion Cycle 3 of 3 Calculate hydrodynamics completed BEST Model classifier Close Stop 100% Help Config

Layout and features of US-SOMO, integrated in UltraScan and available for Linux, Win, and Mac - 0 X SOMO Solution Modeler Lookup Tables SOMO MD PDB Configuration File 1 will be included SOMO Grid Function Options (AtoB) - D × eted SOMO Grid Function Options (AtoB): Computations Relative to: – Center of Mass Center of Cubelet 1 is model Ŧ Cube Side (Angstrom): Ŧ 5 Add theoretical hydration (PDB only) 2 Apply Cubic Grid re popping Adjust Overlap Options Expand Beads to Tangency Enable ASA screening ☑ Help Close Finished with popping and radial reduction Rechecking beads View ASA Results Visualize Bead Model 0 previously buried beads are exposed by rechecking Finished rechecking beads Batch Mode/Cluster Operation View Bead Model File Build bead model completed Load Single Bead Model File 8RAT 1 All options set to default values SAXS/SANS Functions Automatically Calculate Hydrodynamics Begin hydrodynamic calculations Hydrodynamic Calculations: Model 1 will be included Calculate RB Hydrodynamics SMI Calculate RB Hydrodynamics ZENO Processing model 1 bead count 246 vbar 0.71 Using 98 beads for the matrix Show Hydrodynamic Calculations Open Hydrodynamic Calculations File Supermatrix inversion Cycle 1 of 3 Supermatrix inversion Cycle 2 of 3 Select Parameters to be Saved Save parameters to file Supermatrix inversion Cycle 3 of 3 Calculate hydrodynamics completed BEST Model classifier Close Stop 100% Help Config

lease Somo Hydrodynamic Results					
SOMO Hydrodynamic Results (Water at 20°C):					
Total Beads in Model:			246		
Used Beads in Model:			98		
Molecular Mass:			1.3681e+04 Da		
Part. Specif. Volume:			0.710 cm^3/g		
s(20,w):			1.93e+00 S		
D(20,w), transl.:			1.18e-06 cm/sec^2		
Stokes Radius:			1.81e+00 nm		
Radius of Gyration:			1.48e+00 nm		
Relaxation Time, tau(h):			7.81e+00 ns		
Intrinsic Viscosity:			3.24c+00 cm^3/g		
Load ASA Results File			View Bead Model File		
Load Full Hydrodynamics Results File					
Help			Close		
Hydrodynamic Calculations:					
Calculate RB Hydrodynamics SMI			Calculate RB Hydrod	dy.	
Show Hydrodynamic Calculations			Open Hydrodynamic	C	
Select Parameters to be Saved			Save parameters to fil	le	
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SAXS/SANS Functions				
Run	DMD			
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Bead Model Functions:				
Bead Model Suffix:		A	20R50hiC	
Overwrite exis	ting filename	s	Add auto-gen	erated su
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Other hydrodynamic computations methods

ZENO – Developed by Mansfield, Kang and Douglas (Stevens Institute and NIST)

BEST – Developed by S. Aragon, SFSU, CA

Scientific Principle of Program:

The Zeno computational method involves enclosing an arbitraryshaped probed object within a sphere and launching random walks from this sphere. The probing trajectories either hit or return to the launch surface ('loss') as shown in the figure for a model soot particle aggregate, whereupon the trajectory is either terminated or reinitiated.



The fraction of random walk trajectories that hit the probed object determines its capacity C (hydrodynamic radius) and the electric polarizibility tensor α and [η] are estimated similarly.

ZENO: A Monte Carlo Numerical Path Integrator



 $\beta = \frac{\# hits}{\# attempts}$ $R_{H} = \beta R$ $fr = 6\pi \eta R_{H}$ $D = \frac{kT}{6\pi \eta R_{H}}$

Escaping path

Douglas & Zhou & Hubbard, *PRE*, Vol 49, Page 5319, (1994).

Douglas & Garboczi, Adv. Chem. Phys, Vol 91, Pages 85-153,(1995).

Mansfield et al., *PRE*, Vol 53, Vol 64, Pages 061401-16, 2001

ZENO computes:

electric Polarizability Tensor,
self-capacity,
intrinsic conductivity
Intrinsic viscosity
hydrodynamic radius
translational diffusion coefficient
translational friction coefficient
radius of gyration
structure factor...
of arbitrarily shaped objects

http://web.stevens.edu/zeno/





BEST uses a very precise boundary element numerical solution of the exact formulation of the hydrodynamic resistance problem with stick boundary conditions to compute the full transport tensors in the center of resistance or the center of diffusion for an arbitrarily shaped rigid body, including rotation-translation coupling.

The input for BEST is a triangulation of the solvent-defined surface of the molecule of interest, given by Connolly's MSROLL. The triangulation is prepared for BEST by COALESCE, a program that allows user control over the quality and number of triangles to describe the surface. The computations are repeated for a series of triangulated structures with different number of plates, and extrapolated to zero plate size.

How to reliably test the methods?

- We chose to investigate in depth only the translational friction data, namely $D^0_{t(20,w)}$ and $s^0_{(20,w)}$.
- We have collected literature data, and carefully analyzed them for proper extrapolation and reduction to standard conditions.
- We selected PDB crystal structures from the same species as the experimental data.
- We performed the computations on those structures using SoMo, SoMo+Zeno, AtoB with two different grid sizes, BEST (all using the US-SOMO interface), and HYDROPRO (externally).

Test proteins used

#	Monomeric proteins	MW	#	Multimeric proteins	MW
1	Cytochrome c (1HRC)	12357.5	14	Superoxide dismutase (2SOD)	31442.2
2	Ribonuclease A (8RAT)	13683.8	15	β-Lactoglobulin (1BEB)	35224.7
3	α-Lactalbumin (1A4V+carb)	15784.7	16	α-Chymotrypsin (4CHA)	50473.5
4	Lysozyme (1AKI)	14306.7	17	Triosephosph. Isom. (1YPI)	52971.4
5	Myoglobin horse CO (1DWR)	17568.3	18	Hemoglobin CO (1HCO)	64559.7
6	Soybean Trypsin Inh. (1AVU)	19962.8	19	Citrate Synthase (1CTS)	97845.5
7	β-Trypsin (1TPO)	23335.9	20	Inorganic Pyrophosph. (1FAJ)	117339.0
8	Trypsinogen (1TGN)	23182.7	21	G3PD apo (2GD1)	143787.8
9	α-Chymotrypsin (4CHA)	25236.5	22	G3PD holo (1GD1)	146437.7
10	Chymotrypsinogen A (2CGA)	25659.0	23	LDH pig H + NAD (5LDH)	148942.6
11	Carbonic Anhydr. B (2CAB)	28820.5	24	LDH pig M + NAD (9LDH)	149063.5
12	Pepsin (4PEP)	34588.6	25	Aldolase (1ADO)	157136.0
13	H. Serum Albumin (1AO6)	66428.6	26	Catalase (4BLC)	235782.0
			27	β-Galactosidase (1BGL)	465557.0

Overall performance of the different hydrodynamic modeling methods: *D_t*



Comparison between hydrodynamic calculations methods using 21 test proteins: translational diffusion coefficient



Overall performance of the different hydrodynamic modeling methods: s



Comparison conclusions:

- *D_t* is always better matched than *s*. This is likely due to poor psv knowledge/estimation.
- HYDROPRO and BEST both underestimate D_t and s. This is likely due to an excessive expansion of the surface in an attempt to account for hydration.
- SoMo with overlap removal overestimates D_t and s. This is likely due to an excessive shrinkage of the hydrated beads notwithstanding the outward translation.
- AtoB with a 5 Å grid appears to produce reasonable hydrated surfaces leading to very good D_t matching.
- The combination of SoMo models without overlap removal and Zeno computations produces the best D_t matching.